A molecular timescale for vertebrate evolution

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A timescale is necessary for estimating rates of molecular and morphological change in organisms and for interpreting patterns of macroevolution and biogeography1,2. Traditionally, these times have been obtained from the fossil record, where the earliest representatives of two lineages establish a minimum time of divergence of these lineages3. The clock-like accumulation of sequence differences in some genes provides an alternative method4 by which the mean divergence time can be estimated. Estimates from single genes may have large statistical errors, but multiple genes can be studied to obtain a more reliable estimate of divergence time5,6,7. However, until recently, the number of genes available for estimation of divergence time has been limited. Here we present divergence-time estimates for mammalian orders and major lineages of vertebrates, from an analysis of 658 nuclear genes. The molecular times agree with most early (Palaeozoic) and major lineages of vertebrates, from an analysis of 658 nuclear genes. The molecular times agree with most early (Palaeozoic) and late (Cenozoic) fossil-based times, but indicate major gaps in the Mesozoic fossil record. At least five lineages of placental mammals arose more than 100 million years ago, and most of the modern orders seem to have diversified before the Cretaceous/Tertiary extinction of the dinosaurs.

Figure 1 Estimation of divergence times. a, Calibration. The arrow marks the first appearance of synapsids (ancestors of mammals) and diapsids (ancestors of birds) in the fossil record at 310 Myr ago. Reconstructions of an early synapsid (Varanosaurus) and stem diapsid (Hylonomus) are shown. The dark shading represents the reptilian portion and the lighter shading represents the avian and mammalian portion of the phylogeny. b, Two methods for dating the unknown divergence time (t) between A1 and A2 when A and B diverged at calibration time T (Myr ago). In the average distance method, t = d(\frac{1}{2}d(\frac{1}{2})^2), where d = (d_{AB} + d_{BA})/4T is the rate of change for lineages A and B, and d is the number of substitutions per site between sequences i and j. In the lineage-specific method, t_{AB} = d_{ij}/2r_{ij} (where r_{ij} = e_{ij}T), alternatively, t is based on the length of one of the two lineages; that is, t = e_{i} or t = e_{j}. Where e_{i} and e_{j} are estimated by the ordinary least squares method (C = outgroup).

Molecular clocks are first calibrated with a known time of divergence and then used to estimate divergence times of other species. The divergence of birds and mammals provides a reliable calibration point with which to anchor molecular clocks. The earliest ancestors of mammals (synapsids) and birds (diapsids) are lizard-like and first appear in the Carboniferous period, at ~310 million years (Myr) ago8,9 (Fig. 1a). The fact that the fossil record10,11 documents a morphological transition from lobe-finned fishes to stem tetrapods at 370–360 Myr ago, and records the appearance of stem amphibians at 338 Myr ago, indicates that the time of the diapsid–synapsid split (within amniotes) is unlikely to be a considerable underestimate. Alternatively, multiple calibration points based on the mammalian fossil record may be used, but this might result in substantial underestimates of divergence time2,12,13.

We used 658 genes, representing 207 vertebrate species, to estimate divergence times by two methods (see also Fig. 1b; Methods; Supplementary Information). Taxonomic biases in the sequence databases resulted in a predominance of mammalian sequences studied. For estimates derived from large numbers of genes, distributions of divergence-time estimates are approximately normal (Fig. 2a–i; Methods). These distributions show considerable dispersion around the peak, as reflected in high coefficients of variation. On average, standard errors of ~10% were obtained with 10 genes, 5% with 50 genes, and 3% with 100 genes. Multigene time estimates obtained without rate-constancy tests were nearly identical to those obtained with stringent rate testing (Fig. 2j–l). This indicates that there are probably no underlying directional biases in the data.

Molecular times for the origin of the major lineages of vertebrates in the Palaeozoic and early Mesozoic eras are similar to those that are based on the fossil record14 (Fig. 3). The molecular time estimate for the marsupial–placental split, 173 Myr ago, corresponds well with the fossil-based estimate (178–143 Myr ago)15. The bird–crocodilian divergence is slightly younger than the earliest fossils suggest16, at 240 Myr ago, but this difference is less than one standard error. The molecular estimate of the lissamphibian–amniote divergence at 360 Myr ago also agrees with the fossil-based estimate10,11. Fewer genes are available to time the earliest divergences among vertebrates, but molecular times (of 564 and 528 Myr ago) are consistent with the Late Cambrian fossil record for the first appearance of vertebrates (at 514 Myr ago)14.

A striking pattern revealed by our molecular divergence times is the Cretaceous origin of all modern orders of mammals examined (Fig. 3). Earlier molecular12 and fossil15 studies found evidence that at least some mammalian divergences occurred in the Cretaceous, leaving open the possibility of a gradual diversification of orders into the early Cenozoic. Molecular times now indicate that at least five major lineages of placental mammals (Edentata, Hystrixcognathi, Sciuromorpha, Paenungulata, Archonta + Formicata) may have arisen in the Early Cretaceous, >100 Myr ago, and that most mammalian orders were involved in a Cretaceous radiation that predated the Cretaceous/Tertiary extinction of the dinosaurs (Fig. 3). The origin of most mammalian orders seems not to be tied to the filling of niches left vacant by dinosaurs, but is more likely to be related to events in Earth history17. Similarly, a mid-Cretaceous divergence was obtained for the bird orders Anseriformes and Galliformes12,13.

Multigene divergence times within several orders of mammals compare closely with fossil-based estimates. For example, molecular divergence times from humans to chimpanzees, gorillas, gibbons, and Old World monkeys are close to currently accepted dates from the fossil record4,10,18. The orangutan molecular divergence
time (8.2 Myr ago) corresponds with the age of the unique skull of the fossil hominoid *Sivapithecus*<sup>6</sup>, which is usually placed on the orangutan lineage, but is about 4 Myr younger than the earliest teeth and jaw fragments assigned to *Sivapithecus*<sup>6</sup>. The *Sivapithecus*–orangutan association itself has been questioned<sup>20,21</sup>. Molecular time estimates among cetartiodactyls (whales and artiodactyls) and for the catarrhine–platyrrhine and feliform–caniform divergences are close to, or only slightly older than, fossil-based estimates.

In contrast, molecular divergence times among sciurognath rodents (Fig. 3) are roughly four times older than their fossil-based estimates<sup>2</sup>, as was found previously<sup>12,23</sup>. Because these times were estimated from many genes (343) and did not change when a lineage-specific method (Fig. 1b) was used (e.g., a divergence time of 41 Myr ago was obtained for the mouse–rat divergence), the difference cannot be attributed to stochastic error or increased rate of substitution in rodents. Furthermore, increased stringency of the rate-constancy test resulted in similar time estimates (Fig. 2j–l). Overall, fossil-based and molecular times are in relatively close agreement (Fig. 4a), except for the origin of placental orders and the early history of rodents. The average difference between molecular and fossil-based dates for Mesozoic comparisons is large (30%)

**Methods**

Sequence retrieval and tests of molecular clocks. Amino-acid sequences of nuclear genes were obtained from the HOVERGEN<sup>6</sup> database (Genbank Release 97) and all 5,850 gene families were manually examined. Alignments were retrieved whenever data were available to time at least one divergence. Genes under strong positive selection (for example, major histocompatibility complex genes) and sequences with ambiguous orthologies and extensive alignment gaps were excluded. Gene phylogenies were scrutinized further and genes (or sequences) showing extensive rate variation among lineages forms are known. Recent findings of 85-Myr-old placental fossils from Central Asia<sup>13–16</sup> are also evidence for the presence of mammalian fossils in this Mesozoic gap.

Our molecular timescale for vertebrate evolution will be useful in calibrating local molecular clocks and in estimating intraordinal divergence times more reliably, especially in groups with poor fossil records. Molecular times also provide an independent measure of the tempo and mode of morphological change. For example, the sudden appearance (in the Early Tertiary fossil record) of mammalian and avian orders, which show large morphological differences, has been taken to imply rapid rates of morphological change at that time<sup>14,24</sup>. Now, the possibility of 20–70 Myr of prior evolutionary history relaxes that assumption and suggests a greater role for Earth history in the evolution of terrestrial vertebrates<sup>13,15</sup>. An accurate knowledge of divergence times can help to direct the search for ‘missing’ fossils and test hypotheses of macroevolution.

**Figure 2** Histograms (a–i) of distributions of single-gene divergence times for nine multigene time estimates, and graphs (j–l) of the effects of increased stringency of the rate-constancy test (corresponding to areas of 5% (rejection curve) for the same divergences. Divergence of: a, Hominioidea and Cercopithecoida; b, Muridae and Cricetidae; c, Archonta and Ferungulata; d, Ruminantia and Suidae; e, Carnivora+Perissodactyla and Cetartiodactyla; f, Rodentia and (Archonta+Ferungulata+Paenungulata); g, mouse and rat; h, Primates and Lagomorpha; and i, Amphibia and Amniota. j, Percentage of pairwise comparisons not rejected; k, Percentage of genes not rejected; l, Time estimates for divergences a–i. Symbols for histograms: *M*, mode; *m*, mean; *N*, total number of genes; *n*, number of genes used after removal of outliers; *V*, coefficient of variation. The locations of *m* and *M* are shown. Myr, millions of years ago.
Figure 3 A molecular timescale for vertebrate evolution. All times indicate Myr separating humans (or the largest sister group containing humans) and the group shown, except when the comparative groups are separated by a slash (/). Time estimates are shown with ±1 s.e.m. and the number of genes used is given in parentheses. Three groups of mammalian orders are: Archonta (Primates, Scandentia, Dermoptera, Chirotteria, and Lagomorpha), Ferungulata (Carnivora, Carnivora/Perissodactyla), and Paenungulata (Hyracoidea, Proboscidea, Sirenia). Cam, Cambrian; Carbonif, Carboniferous; Dev, Devonian; PC, Precambrian; Perm, Permian; Prot, Proterozoic; Sili, Silurian; Tri, Triassic.

Figure 4 Comparison of fossil-based and molecular estimates of divergence time in vertebrates. a, Plot of all values (correlation coefficient = 99%). The solid line indicates a 1:1 relationship; the closed circle represents the calibration point. b, Close-up of the region from 0 to 150 Myr ago. Open diamonds show fossil dates and the solid line shows divergence time estimated using the molecular clock.

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Letters to Nature

The ParaHox gene cluster is an evolutionary sister of the Hox gene cluster

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Genes of the Hox cluster are restricted to the animal kingdom and play a central role in axial patterning in divergent animal phyla. Despite its evolutionary and developmental significance, the origin of the Hox gene cluster is obscure. The consensus is that a primordial Hox cluster arose by tandem gene duplication close to animal origins. Several homeobox genes with high sequence identity to Hox genes are found outside the Hox cluster and are known as ‘dispersed’ Hox-like genes; these genes may have been transposed away from an expanding cluster. Here we show that three of these dispersed homeobox genes form a novel gene cluster in the cephalochordate amphioxus. We argue that this ‘ParaHox’ gene cluster is an ancient parologue (evolutionary sister) of the Hox gene cluster; the two gene clusters arose by duplication of a ProtoHox gene cluster. Furthermore, we show that amphioxus ParaHox genes have co-linear developmental expression patterns in anterior, middle and posterior tissues. We propose that the origin of distinct Hox and ParaHox genes by gene-cluster duplication facilitated an increase in body complexity during the Cambrian explosion.

Homeodomain sequence comparisons reveal that at least five classes of homeobox genes are as closely related to Hox genes as many of the latter are to each other. These are the Evx, Mox, Cdx (or cad), Xlox, and Gsx homeobox classes (we term a class defined by mouse Gsh-1 and Gsh-2 as Gsx). The two mammalian Evx genes are each linked to the 5' end of Hox gene clusters, and a cnidian Evx-like gene is linked to a Hox-like gene, indicating that the close sequence relationship between Evx and Hox genes reflects tandem duplication. Mox genes may represent a similar case because the mouse Mox-1 gene maps to chromosome 11, close to the Hoxb cluster. The Cdx, Xlox and Gsx gene families are more problematic.

To investigate the evolutionary origins of Cdx, Xlox and Gsx genes, we elected to clone representatives of each gene family from amphioxus. This is because homeobox gene families in this animal are not complicated by either excessive duplication (as in vertebrates) or divergence and rearrangement (as in Drosophila or nematode). Using primers directed to Hox class homeoboxes and amphioxus genomic DNA as template, amplification by polymerase chain reaction (PCR) yielded partial clones of Cdx and Xlox class homeoboxes. A fragment of amphioxus Gsx was also cloned by PCR, using primers designed from the two mammalian gene family members Gsh-1 and Gsh-2. To determine the complete homeobox sequence of each gene, we isolated longer clones from amphioxus genomic libraries: only single members of each class were obtained, which we named AmphiCdx, AmphiXlox and AmphiGsx. Their encoded homeodomains resemble those of the Drosophila or vertebrate homologues (Fig. 1).

Analysis of genomic clones revealed that amphioxus Xlox and Cdx class homeoboxes were unexpectedly contained within a single bacteriophage clone. Mapping indicated that the homeoboxes were separated by just 7.5 kilobases (kb). Furthermore, using genomic walking we found that these two homeobox genes are physically linked to the AmphiGsx gene. The Gsx and Xlox homeoboxes are separated by just 25 kb (Fig. 2). We designate this tight cluster of three genes the ParaHox gene cluster.

The finding that amphioxus Gsx, Xlox and Cdx class genes form a novel homeobox cluster challenges the idea that these homeobox gene clusters are ‘dispersed’ Hox genes. To reconcile linkage in